INTRODUCTION

U-74389G is not famous for its hypazotemic capacity ($P = 0.0005$). U-74389G, as a novel antioxidant factor, implicates exactly only 256 published studies. The hypoxia-reoxygenation (HR) type of experiments was noted in 4.29% of these studies. A tissue-protective feature of U-74389G was obvious in these HR studies. The U-74389G chemically known as 21-[4-(2,6-di-1-pyrrolidinyl-4-pyrimidinyl]-1-piperazinyl]-pregna-1,4,9(11)-triene-3,20-dione maleate salt is an antioxidant complex, which prevents the lipid peroxidation either iron-dependent or arachidonic acid-induced one. Animal kidney, liver, brain microvascular endothelial cells monolayers, and heart models were protected by U-74389G administration. E-74389G also attenuates leukocytes, downregulates the proinflammatory gene, treats the endotoxin shock, produces cytokine, enhances the mononuclear immunity, protects the endothelium, and presents antishock property.

Erythropoietin (Epo) even whether is not famous for its hypazotemic action ($P = 0.4430$), it can be used as a reference.
drug for comparison with U-74389G. Although Epo is met in over 30,022 published biomedical studies, only a 10.53% of them negotiate the known type of HR experiments. Nevertheless, Epo as a cytokine, it is worth being studied about urea levels too.

This experimental work tried to compare the hypazotemic effects of the above drugs on a rat-induced HR protocol. They were tested by calculating the serum urea levels declines.

**MATERIALS AND METHODS**

**Animal preparation**
The Vet licenses between 3693/12-11-2010 and 14/10-1-2012 numbers, the granting company and the experiment location are mentioned in preliminary references.[1,2]The human animal care of albino female Wistar rats, the 7 days preexperimental ad libitum diet, the nonstop intraexperimental anesthesiologic techniques, the acidometry, the electrocardiogram and the oxygen supply, and postexperimental euthanasia are also described in preliminary references. Rats were 16–18 weeks old. They were randomly assigned to six groups consisted of \( N = 10 \). The stage of 45 min hypoxia was common for all six groups. Afterwards, reoxygenation of 60 min was followed in group A; reoxygenation of 120 min in group B; immediate Epo intravenous (IV) administration and reoxygenation of 120 min in group C, immediate Epo IV administration and reoxygenation of 120 min in group D, immediate U-74389G IV administration and reoxygenation of 60 min in group E, and immediate U-74389G IV administration and reoxygenation of 120 min in group F. The dose height assessment for both drugs is described at preliminary studies as 10 mg/kg body mass.

Hypoxia was caused by laparotomic clamping the inferior aorta over renal arteries with forceps for 45 min. The clamp removal was restoring the inferior aorta patency and reoxygenation. After exclusion of the blood flow, the protocol of HR was applied, as described above for each experimental group. The drugs were administered at the time of reperfusion through inferior vena cava catheter. The urea levels were determined at 60\(^{th}\) min of reoxygenation (for A, C, and E groups) and at 120\(^{th}\) min of reoxygenation (for B, D, and F groups). However, the predicted urea values were used since a very powerful relation was raised with animals’ mass \( (P = 0.0111) \).

**Statistical analysis**
Table 1 presents the (%) hypazotemic influence of Epo regarding reoxygenation time. Furthermore, Table 2 presents the (%) hypazotemic influence of U-74389G regarding reoxygenation time. Chi-square tests were applied using the ratios which produced the (%) results per endpoint. The outcomes of Chi-square tests are depicted in Table 3. The statistical analysis was performed by Stata 6.0 software (Stata 6.0, StataCorp LP, Texas, USA).

**RESULTS**
The successive application of Chi-square tests revealed that U-74389G accentuated the hypazotemia by 158.4209-fold \( (157.8723–158.9713) \) than Epo at 1 h, by 4.50889-fold \( (4.506346–4.511436) \) at 1.5 h, by 2.850291-fold \( (2.846433–2.854153) \) at 2 h, by 0.9017775-fold \( (0.8996649–0.903895) \) without drugs, and by 4.632148-fold \( (4.628636–4.635662) \) whether all variables have been considered \( (P = 0.0000) \).

**DISCUSSION**
The unique available study investigating the hypazotemic effect of U-74389G on urea levels was the preliminary one.[1] Although the most famous activities of neuroprotection and membrane-stabilization properties, it accumulates the cell membrane, protecting vascular endothelium from peroxidative damage but hardly penetrates the blood–brain barrier. It elicits a beneficial effect in ototoxicity and Duchenne muscular dystrophy. It increases γ-glutamyltransferase, superoxide dismutase (SOD), and glutathione levels in oxygen-exposed cells. It treats septic states and acts as an immunosuppressant in flap survival. It prevents the learning impairments, and it delays the early synaptic transmission decay during hypoxia improving the energetic state of...
neurons. It shows antiproliferative properties on brain cancer cells and is considered as a new promising anti-inflammatory drug for the treatment of reperfusion syndrome in IR injuries.

The same authors confirmed the short-term hypoxiaemic effect of Epo preparations in noniron-deficient individuals. Zeng et al. noticed that blood urea nitrogen (BUN) was decreased by cyclic helical B-peptide derived from Epo in a dose-dependent fashion maybe through reducing caspase-3 activation, apoptosis, and inflammation in a mice model. Ma et al. levels showed small but significant improvement of BUN than control treatment after the delivery of human Epo cDNA to salivary glands. Tajbakhsh et al. the postdialysis means of malondialdehyde, BUN, advanced glycation end product (AGE), and ox-low-density lipoprotein plasma level significantly decreased than predialysis level in chronic hemodialysis end-stage renal disease patients on maintenance treatment with Epo therapy. Saraji et al. found no significant relation between restless leg syndrome and dialysis adequacy, serum intact parathyroid hormone, and urea; whereas, Epo dosage and serum hemoglobin level were lower. Koriem et al. found significant increases in serum Epo and urea levels in 4-tert-cyclophenoxyl toxicity – an endocrine-disrupting chemical in rats. Yamazaki et al. found no significant differences regarding the dose of darbepoetin and BUN between transplant and no transplant chronic kidney disease (CKD) dialysis recipients. Huang et al. found high blood lead level ( BLL ) 0.318-fold rarely in the group with Epo than in the group without Epo. Clearance of urea was positively associated with log BLL in maintenance hemodialysis patients. Tahamtan et al. could not reverse the increased level of BUN by Epo 24 h after reperfusion, but counteracted 7 days after bilateral renal ischemia in Wistar rats. Hertzberg-Bigelman et al. reflected the elevated BUN, presence of anemia, and molecular changes in the Epo/Epo-R axis in a severe renal failure rat model. Schaal et al. found a significant positive correlation between antimicrobial β-defensin-like role of hepcidin (HEPC) levels and BUN by 1.8-fold and decreased Epo levels by 77.8% in septic acute kidney injury (AKI) patients before and after treatment. Maduell et al. significantly decreased blood urea nitrogen values increasing the dialysis dose; whereas, there was a 50.4% of reduction in darbepoetin dose at 24 months and a significant decrease in the Epo resistance index in online hemodiafiltration. Antwi-Bafour et al. observed a significantly increased fasting blood urea by 1.45-fold and Epo levels also high as 54.0% in the cases than controls in renal insufficiency patients with poorly controlled diabetes. Xu et al. found no obvious changes of BUN and Epo in either groups after sodium ferrous chlorophyll treatment of hemodialysis patients; whereas, Epo dosage was decreased more significantly than in the control group (P < 0.05). Vega et al. found the mean equivalent renal urea clearance at 15.6 ± 4.2 mL/min and the Epo resistance index at 3.7 ± 2.1 IU/kg/week/gDL in patients who received home hemodialysis. Ibrahim et al. exhibited reduced serum levels of Epo and increases in the kidney function markers (urea and creatinine) in male Wistar rats of induced diabetes mellitus than control ones. Xue et al. showed a significantly increasing similar trend (P < 0.05) for both Epo levels and BUN postoperatively in a rat model of chronic allograft nephropathy. Elshiekh et al. decreased urea contents in the IR kidney after treatment with Epo or ischemic preconditioning (IPC). Liu et al. significantly decreased serum expression levels of BUN markedly induced by renal ischemia-reperfusion injury after pretreatment with Epo and sodium selenite. Hirata et al. alleviated the increasing urinary total protein excretion and plasma urea nitrogen, in a rat model of chronic progressive glomerulonephritis (GN), thereby delaying end-stage kidney disease after a single dose of epoetin beta pegol (continuous Epo receptor activator [CERA] given on day 1). Cakirgolu et al. improved statistically nonsignificantly the serum urea particularly after daily Epo application in AKI rats. Phillips et al. suggested inefficient erythropoiesis in the presence of elevated Epo that is correlated with plasma urea concentration in Lewis polycystic kidney CKD rodent model. Zhao et al. calculated BUN levels less severe after administration of Epo at a dose of 3000 IU/kg once a week than the placebo group at day 21 (P < 0.01) in rats model of induced renal interstitial fibrosis. Banks conducted isothermal urea denaturation experiments at numerous temperatures in the absence and presence of sodium chloride and indicated that salt stabilizes Epo primarily by increasing the difference in enthalpy between the native and unfolded states. This result, and the finding that the salt induced increases in Epo, melting temperatures were independent of the identity of the salt cation, and anion indicates that salt likely increases the conformational stability of Epo at neutral pH by nonspecific shielding of unfavorable electrostatic interaction(s) in the native state. Attia et al. related high predialysis levels of BUN with uremic pruritus, a common treatment of which includes also Epo in dialysis patients with chronic kidney disease. Bueno et al. noticed increase in median Epo levels (P < 0.001) and urea (P = 0.039) in the evaluated period in chronic kidney disease patients. Yang et al. found that nandrolone decanoate plus Epo may increase HCT by 2.54%; whereas, no evidence was found to indicate that decreased BUN than Epo alone. Reddy et al. documented Epo dosage and urea reduction rate in maintenance hemodialysis patients per week. Dardashi et al. found statistically no significant differences in p-urea and IV administration of a single high-dose (400 IU/ kg) Epo, without renal protective effect in patients with reduced kidney function undergoing coronary artery bypass surgery. Aydin et al. positively correlated prohepcidin with urea (P = 0.002) levels; whereas, HEPC generation is also dependent on Epo levels in chronic kidney disease. Zhang et al. maintained for < 1 week, the declines in Hb and BUN following the end of the treatment with chitosan-tripolyphosphate-Epo nanoparticles in an immunoglobulin A nephropathy (IgAN) rat model. Aizawa et al. increased urea nitrogen after epoetin beta pegol (CERA) in Thy-1-GN rats. Arend et al. considered the dose-dependent adverse effects of high darbepoetin treatment as elevated serum urea in nephrectomized apo E knockout mice. Ahmadian et al. significantly decreased blood SOD, GPx,
and urea levels and increased TAC level after treatment with Epo and MEL in renal IR injury of male rats. Nasri found that Epo was able to prevent the increase in serum BUN. Thus, Epo is a promising kidney protective agent to prevent, ameliorate, or attenuate tubular damage induced by nephrotoxic agents in a study of male rats. Siribamrungwong et al. improved the nutritional status since BUN was increased by 1.09% and Epo dosage requirement was decreased by 25%/week in peritoneal dialysis patients. Chrysikos DT et al. found serum urea not to differ between the two groups of a swine model of pancreatic IR encompassing IPC and U-74389G effect. Alhan et al. inhibited the significant increase in serum urea after the use of U-74389G in induced acute necrotizing pancreatitis in rats. According to above, Table 3 shows that U-74389G accentuated by 4.632148-fold (4.628636–4.635662) the hypozotemic potency than Epo (P = 0.0000); a trend attenuated along time, in Epo nondeficient rats. A meta-analysis of these ratios from the same experiment, for 12 other seric variables, provides comparable results [40] [Table 4].

CONCLUSION

The antioxidant agent U-74389G proved also more hypozotemic by 4.632148-fold (4.628636–4.635662) than Epo (P = 0.0000) in rats. However, this trend is attenuated along the short-term time frame of the experiment. A biochemical investigation is needed for the elucidation of this metabolic pathway.

ACKNOWLEDGMENT

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Table 3: The U‑74389G/erythropoietin efficacies ratios on urea levels hypozotemia after Chi-square tests application

<table>
<thead>
<tr>
<th>Odds ratio</th>
<th>95% Confidence interval</th>
<th>P values</th>
<th>Endpoint</th>
</tr>
</thead>
<tbody>
<tr>
<td>158.4209</td>
<td>157.8723–158.9713</td>
<td>0.0000</td>
<td>1 h</td>
</tr>
<tr>
<td>4.50889</td>
<td>4.506346–4.511436</td>
<td>0.0000</td>
<td>1.5 h</td>
</tr>
<tr>
<td>2.850291</td>
<td>2.846433–2.854153</td>
<td>0.0000</td>
<td>2 h</td>
</tr>
<tr>
<td>0.9017775</td>
<td>0.8996694–0.903895</td>
<td>0.0000</td>
<td>Reperfusion time</td>
</tr>
<tr>
<td>4.632148</td>
<td>4.628636–4.635662</td>
<td>0.0000</td>
<td>Interaction</td>
</tr>
</tbody>
</table>

Table 4: A U‑74389G/erythropoietin efficacies ratios meta‑analysis on 12 hematologic variables (10 variables with balancing efficacies and two variables with opposite efficacies) [40]

<table>
<thead>
<tr>
<th>Variable</th>
<th>1 h</th>
<th>1.5 h</th>
<th>2 h</th>
<th>P value</th>
<th>Reperfusion time</th>
<th>Interaction</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>WBC</td>
<td>0.957451</td>
<td>0.3782</td>
<td>1.396122</td>
<td>0.0000</td>
<td>1.918237</td>
<td>0.0000</td>
<td>1.71622</td>
</tr>
<tr>
<td>Hematocrit</td>
<td>38.424</td>
<td>0.0000</td>
<td>9.076658</td>
<td>0.0000</td>
<td>6.222898</td>
<td>0.0000</td>
<td>1.001356</td>
</tr>
<tr>
<td>Hemoglobin</td>
<td>1.268689</td>
<td>0.0000</td>
<td>1.839035</td>
<td>0.0000</td>
<td>13.1658</td>
<td>0.0000</td>
<td>1.252422</td>
</tr>
<tr>
<td>RBC count</td>
<td>0.961059</td>
<td>0.0000</td>
<td>1.733395</td>
<td>0.0000</td>
<td>6.519657</td>
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<td>RbcDW</td>
<td>3.307733</td>
<td>0.0000</td>
<td>3.023389</td>
<td>0.0000</td>
<td>2.655885</td>
<td>0.0000</td>
<td>0.2259914</td>
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<tr>
<td>Platelet count</td>
<td>2.42839</td>
<td>0.0000</td>
<td>6.00238</td>
<td>0.0000</td>
<td>6.1333429</td>
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<td>3.939027</td>
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<td>MPV</td>
<td>145.8532</td>
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<td>0.0000</td>
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<tr>
<td>Platelet DW</td>
<td>0.6940233</td>
<td>0.0000</td>
<td>1.319118</td>
<td>0.0000</td>
<td>2.206972</td>
<td>0.0000</td>
<td>2.2484006</td>
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<td>Creatinine</td>
<td>168.9034</td>
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<td>4.872332</td>
<td>0.0000</td>
<td>3.039572</td>
<td>0.0000</td>
<td>1.0262016</td>
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<td>Total proteins</td>
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<td>0.0000</td>
<td>4.421079</td>
<td>0.0000</td>
<td>2.803573</td>
<td>0.0000</td>
<td>0.8842162</td>
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<tr>
<td>Mean</td>
<td>7.91129943</td>
<td>0.0378</td>
<td>3.10924261</td>
<td>0.0000</td>
<td>3.90159936</td>
<td>0.0000</td>
<td>1.1749027</td>
</tr>
<tr>
<td>Mean corpuscular hemoglobin concentrations</td>
<td>−0.2774225</td>
<td>0.0000</td>
<td>−0.5504722</td>
<td>0.0000</td>
<td>−0.8522433</td>
<td>0.0000</td>
<td>+3.044774</td>
</tr>
<tr>
<td>Platelet crit</td>
<td>−0.2312044</td>
<td>0.0000</td>
<td>−0.6719365</td>
<td>0.0000</td>
<td>−1.330756</td>
<td>0.0886</td>
<td>5.620077</td>
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<tr>
<td>Mean</td>
<td>−0.2532076</td>
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<td>−0.6081795</td>
<td>0.0000</td>
<td>−1.0649544</td>
<td>0.0443</td>
<td>4.1366488</td>
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Greece. The research facilities for this project were provided by the aforementioned institution.

REFERENCES


